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# An assessment of Blue Carbon sources and potential in the Nisqually Estuary using biomarkers and compound-specific isotopes of marsh plants, eelgrass, and sediment

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An assessment of Coastal Blue Carbon sources and potential in the Nisqually Estuary using biomarkers and compound-specific isotopes of marsh plants, eelgrass, and sediment

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#### BACKGROUND

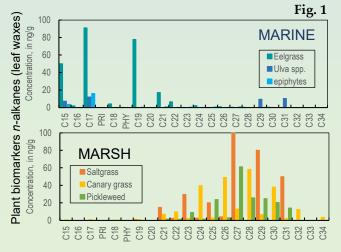
Coastal Blue Carbon ecosystems (e.g., marshes, seagrasses, kelp) sit at the land-sea interface and thus receive organic carbon ( $C_{org}$ ) inputs from adjacent ecosystems. Differentiating between in situ versus transported  $C_{org}$  sources is fundamental to avoiding duplication in Coastal Blue Carbon accounting.

#### STUDY GOALS

Develop a suite of distinctive geochemical tracers, or signatures, that can differentiate among various  $C_{org}$  sources to advance Blue Carbon budgets.

#### APPROACH

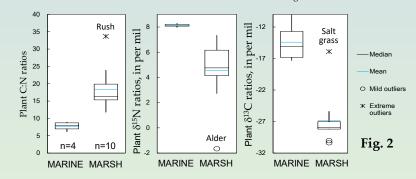
Tidal flow restoration in the Nisqually River Delta is altering land to sea fluxes of terrestrial, marsh, and marine  $C_{org}$  and sediment. Plant end members were characterized to develop multivariate  $C_{org}$  source signatures, which were then applied in a nearshore sediment core to determine  $C_{org}$  sourcing over time.





## **RESULTS – BIOMARKERS and ISOTOPES**

 $C_{org}$  from marine and marsh plants had **distinct** *n*-alkane distributions (Fig. 1), C:N ratios,  $\delta^{13}C_{n-alk}$ ,  $\delta^{15}N$  ratios, and distinct  $\delta^{13}C$  ratios except for salt grass, which had a marine-like  $\delta^{13}C$  ratio (Fig. 2). Sterols did not distinguish between marine and terrestrial  $C_{org}$  sources (not shown).



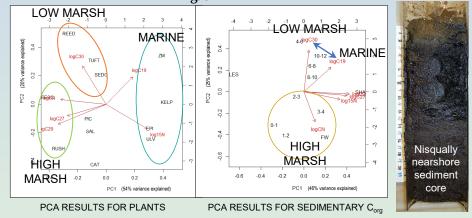
#### PRINCIPAL COMPONENT ANALYSIS (PCA)

Multivariate  $C_{org}$  source signatures were determined by PCA on log-transformed C:N ratios,  $\delta^{15}N$ , and select *n*-alkanes ( $C_{19}$ ,  $C_{27}$ ,  $C_{29}$ ,  $C_{30}$ ) in marine and marsh plants. The same variables were used to reconstruct  $C_{org}$  sourcing by PCA in marsh sediment (LES, SHA, FW) and the sediment record of a nearby eelgrass bed.  $\delta^{13}C$  data is skewed and cannot be used as a variable. PCA was performed using the 'prcomp' function in R software.

ANALYTICAL METHODS – Plant biomarker (*n*-alkanes, sterols) extraction and analysis methods are available upon request. Methods for bulk  $\delta^{13}C$ ,  $\delta^{15}N$ , and C:N ratios in plant and sediment are available from UC Santa Cruz at https://sites.google.com/ucsc.edu/sil; and from UC Davis for CSIA  $\delta^{13}C_{n-alk}$ ,  $\delta^{13}C_{sterol}$  at http://stableisotopefacility.ucdavis.edu.

## PCA RESULTS – PLANTS

- C<sub>org</sub> sources were characterized by (Fig. 3):
- Marine plants:  $\delta^{15}$ N, C<sub>19</sub> *n*-alkane
- Low marsh plants: C<sub>30</sub> *n*-alkane
- High marsh: C:N ratio, C<sub>27</sub>, C<sub>29</sub> *n*-alkanes
- Pickleweed, salt grass, and cattail were poorly characterized by this PCA because no variables plotted in their vicinity.
   Fig. 3



## PCA RESULTS – SEDIMENTS

Two principal components (PCs) explained 71% of the geochemical variance. PC1 describes estuarine sediment with terrestrial  $C_{org}$  (LES, SHA). PC2 describes nearshore sediment with high marsh  $C_{org}$  in the upper 0-4 cm, and mixed low marsh/marine  $C_{org}$  in deeper sediment (4-12 cm) that is stabilized by the presence of eelgrass roots and rhizomes.

## SIGNIFICANCE for BLUE CARBON, RESTORATION

- The restoring Nisqually Estuary affects Coastal Blue Carbon budgets because deeper buried C<sub>org</sub> is from marine and low marsh plants with lower C:N ratios and lower Blue Carbon potential.
- C<sub>org</sub> delivery to nearshore sediments has transitioned to more terrestrial sources (high marsh, not 'blue' carbon) as restoration progresses.