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Appendix S2

To generate a species x trait matrix, we first mined existing data sources (Barnett et al. 2007, Hébert et al. 2016). As these are the most comprehensive databases that we are aware of, we cross-referenced taxa from our lakes with these databases, and eliminated traits that were largely incomplete across most taxa. From these sources, we were able to find comprehensive data for the following: habitat (categorical: pelagic, littoral); feeding mode (categorical: stationary suspension, filter, raptorial, current cruiser); trophic level (categorical: herbivore, omnivore, omnivore-carnivore); body size (continuous); and body mass (continuous). Categorical traits were converted to binary form (0,1) and the trait state littoral was removed because it is perfectly correlated with the pelagic trait state. Because of uncertainties in body size measurements, this variable was converted to ordinal categories using quartiles of the continuous data to generate roughly even numbers of species within each category (in mm, category 1 = ≤ 0.614 , category 2 = $0.615 - 0.880$, category 3 = $0.881 - 1.140$, category 4 = >1.140). To incorporate life history variation, we estimated species-specific egg development rate at 14°C from existing formulae (Table S1). This water temperature was chosen as it represented the average of all lake water temperature profiles throughout the duration of the study. All trait data are summarized in Table S2.

Some categories present in original data sources were pooled here because of small sample sizes. This pooling was done for: 1) B-, C-, D-, and S-filtration species in Barnett et al. (2007) (combined to one category of filter feeders; Table S2); and 2) carnivores and more carnivorous omnivores (OmniCarn in Barnett et al. 2007) (combined to one category omnivore-carnivore; Table S2). If species and specific trait values were not present in existing trait

databases, we took three approaches to fill gaps (denoted in Table S2). First, if values from species within the same genus were present, they were used. Second, we searched the literature for additional values using Google Scholar. Third, we estimated some values with statistical models. The latter was used to estimate body mass for two species using known length-weight regressions (Bottrell et al. 1976). There were three species missing data on several traits. Two of these species, *Kurzia media* and *Oxyurella brevicaudis*, were removed, as they never comprised more than 1% of community abundance and were each present only once in our dataset. The third species, *Aglaodiaptomus leptopus*, was more prevalent; for this species, professional judgement was used to assign a category for feeding mode (stationary suspension), which is consistent with other diaptomid copepods (*Leptodiaptomus*, *Skistodiaptomus*).

Table S1. Model estimates of egg development time (K, days) for different crustacean zooplankton at 14°C water temperature. *a*, *b*, and *c* refer to model coefficients.

Species	Formula/ Source	<i>a</i>	<i>b</i>	<i>c</i>	K (days)
<i>Bosmina</i>	1	3705848	-15.4	-3.11	4.189
<i>Ceriodaphnia</i>	1	1763	3.7	-1.28	3.712
<i>Chydorus</i>	1	138279	-4.9	-2.43	4.558
<i>Diaphanosoma</i>	1	1767	-1.9	-1.08	3.711
<i>Daphnia</i>	1	65912	-6.1	-2.12	4.742
<i>Diaptomus</i>	1	38474	-3.7	-2	5.117
Other cyclopoids	1	7590	-3.4	-1.4	5.798
<i>Tropocyclops</i>	2	18901	-4.8	-1.77	4.375
<i>Diacyclops</i>	2	18901	-4.8	-1.77	4.375
<i>Holopedium</i>	3	10.25638	1.2472	-0.5647	5.400
<i>Polyphemus</i>	3	10.25638	1.2472	-0.5647	5.400
Other cladocerans	3	10.25638	1.2472	-0.5647	5.400
<i>Epischura</i>	4				10.520

- 1) $K = (a(T-b)^c)/24$ (Cooley et al. 1986)
- 2) $K = (a(T-b)^c)/24$ (Borgmann et al. 1984)
- 3) $\ln K = \ln a + b \ln T + c(\ln T)^2$ (Bottrell et al. 1976)
- 4) $K = 0.0791T^2 - 3.7731T + 47.84$ (quadratic equation developed from data in Chen and Folt 1996)

Table S2. Species trait data. Some categories present in original data sources were pooled here because of small sample sizes (see text for details). If species was not present in existing trait databases or had incomplete trait data, either values from species within the same genus were used (indicated with *), literature searches were conducted (see Notes/References), or estimates were derived from statistical models (indicated with †).

Species	Habitat	Feeding	Trophic	Body length (mm)	Body size category	Egg development time (days, at 14 °C)	Body mass (mg)	Notes/References
<i>Acanthocyclops robustus</i>	Pelagic	Raptorial	Omnivore	1.14	3	5.80	0.0086	1,2
<i>Aglaodiaptomus leptopus</i>	Pelagic	Stationary suspension	Omnivore	1.74	4	5.12	0.0198	2, feeding (professional judgement)
<i>Alona setulosa</i> *	Littoral	Filter	Herbivore	0.51	1	5.40	0.0015	1,2
<i>Bosmina longirostris</i>	Pelagic	Filter	Herbivore	0.37	1	4.19	0.0018	1,2
<i>Camptocercus</i> spp.	Littoral	Filter	Herbivore	0.78	2	5.40	0.0051	1,2
<i>Ceriodaphnia lacustris</i>	Pelagic	Filter	Herbivore	0.63	2	3.71	0.0007	1,2
<i>Chydorus</i> spp.*	Littoral	Filter	Herbivore	0.32	1	4.56	0.0030	1,2
<i>Daphnia ambigua</i>	Pelagic	Filter	Herbivore	0.88	2	4.74	0.0139	1,2
<i>Daphnia mendotae</i>	Pelagic	Filter	Herbivore	1.50	4	4.74	0.0332	1,2
<i>Daphnia pulicaria</i>	Pelagic	Filter	Herbivore	1.53	4	4.74	0.0050	1,2
<i>Diacyclops thomasi</i>	Pelagic	Raptorial	Omnivore-carnivore	0.89	3	4.38	0.0032	1,2
<i>Diaphanosoma birgei</i>	Littoral	Filter	Herbivore	0.61	1	3.71	0.0030	1,2
<i>Epischura lacustris</i>	Pelagic	Current-cruiser	Omnivore-carnivore	1.64	4	10.52	0.0173	1,2

Species	Habitat	Feeding	Trophic	Body length (mm)	Body size category	Egg development time (days, at 14 °C)	Body mass (mg)	Notes/References
<i>Holopedium gibberum</i>	Pelagic	Filter	Herbivore	0.86	2	5.40	0.0051	1,2
<i>Latona glacialis</i> *	Littoral	Filter	Herbivore	1.73	4	5.40	0.0050	1,2
<i>Leptodiaptomus novamexicanus</i> *	Pelagic	Stationary suspension	Omnivore	0.96	3	5.12	0.0053	1,2
<i>Leptodiaptomus signicauda</i> *	Pelagic	Stationary suspension	Omnivore	0.96	3	5.12	0.0053	1,2
<i>Leydigia</i> spp.	NA	Filter	Herbivore	0.75	2	5.40	0.0080	1,2
<i>Macrothrix laticornis</i>	NA	Filter	Herbivore	0.47	1	5.40	0.0022	1,2
<i>Mesocyclops edax</i>	Pelagic	Raptorial	Omnivore-carnivore	1.12	3	5.80	0.0117	1,2
<i>Microcyclops varicans</i> *	Littoral	Raptorial	Omnivore	0.61	1	5.80	0.0006†	size (3), size and feeding (4), habitat (5)
<i>Moina macrocopa</i>	Pelagic	Filter	Omnivore	1.14	3	5.40	0.0040†	size (6), trophic (7), habitat (8)
<i>Polyphemus pediculus</i>	Pelagic	Raptorial	Omnivore-carnivore	0.83	2	5.40	0.0029	1,2
<i>Skistodiaptomus oregonensis</i>	Pelagic	Stationary suspension	Omnivore	1.20	4	5.12	0.0078	1,2
<i>Tropocyclops mexicanus</i> *	Pelagic	Raptorial	Omnivore	0.60	1	4.38	0.0015	1,2

1) Barnett et al. (2007); 2) Hébert et al. (2016); 3) Reid (1992); 4) Soto and Hurlbert (1991); 5) Fairchild (1981); 6) Gu et al. (2017); 7) Kumar and Hwang (2008); 8) Santangelo et al. (2018)

References

- Barnett, A. J., K. Finlay, and B. E. Beisner. 2007. Functional diversity of crustacean zooplankton communities: towards a trait-based classification. *Freshwater Biology* **52**:796-813.
- Borgmann, U., H. Shear, and J. Moore. 1984. Zooplankton and potential fish production in Lake Ontario. *Canadian Journal of Fisheries and Aquatic Sciences* **41**:1303-1309.
- Bottrell, H. H., A. Duncan, Z. M. Gliwicz, E. Grygierek, A. Herzig, A. Hillbricht-Ilkowska, H. Kurasawa, P. Larsson, and T. Weglenska. 1976. A review of some problems in zooplankton production studies. *Norwegian Journal of Zoology* **24**:419-456.
- Chen, C. Y., and C. L. Folt. 1996. Consequences of fall warming for zooplankton over wintering success. *Limnology and Oceanography* **41**:1077-1086.
- Cooley, J. M., J. E. Moore, and W. T. Geiling. 1986. Population dynamics, biomass, and production of microzooplankton in the Bay of Quinte during changes in phosphorus loadings. Pages 166-176 in C. K. Minns, D. A. Hurley, and K. H. Nicholls, editors. *Project Quinte: point-source phosphorus control and ecosystem response in the Bay of Quinte, Lake Ontario*. Canadian Special Publications of Fisheries and Aquatic Sciences.
- Fairchild, G. W. 1981. Movement and microdistribution of *Sida crystallina* and other littoral microcrustacea. *Ecology* **62**:1341-1352.
- Gu, L., K. Lyu, Z. Dai, M. Fan, X. Zhu, J. Wang, X. Wang, and Z. Yang. 2017. Predator-specific responses of *Moina macrocopa* to kairomones from different fishes. *International Review of Hydrobiology* **102**:83-89.
- Hébert, M.-P., B. E. Beisner, and R. Maranger. 2016. A compilation of quantitative functional traits for marine and freshwater crustacean zooplankton. *Ecology* **97**:1081-1081.
- Kumar, R., and J. S. Hwang. 2008. Ontogenetic shifts in the ability of the cladoceran, *Moina macrocopa* Straus and *Ceriodaphnia cornuta* Sars to utilize ciliated protists as food source. *International Review of Hydrobiology* **93**:284-296.
- Reid, J. W. 1992. Copepoda (Crustacea) from fresh waters of the Florida Everglades, U.S.A., with a description of *Eucyclops conrowae*. *Transactions of the American Microscopical Society* **111**:229-254.
- Santangelo, J. M., B. N. Soares, T. Paes, P. Maia-Barbosa, R. Tollrian, and R. L. Bozelli. 2018. Effects of vertebrate and invertebrate predators on the life history of *Daphnia similis* and *Moina macrocopa* (Crustacea: Cladocera). *Ann. Limnol. - Int. J. Lim.* **54**:25.
- Soto, D., and S. H. Hurlbert. 1991. Long-term experiments on calanoid-cyclopoid Interactions. *Ecological Monographs* **61**:245-266.